

Uche Eunice Ekpunobi<sup>\*</sup>, Augustine Nnaluo Eboatu, Patrice-Anthony Chudi Okoye and Theresa Uzoma Onuegbu Department of Pure and Industrial Chemistry, Nnamdi Azikiwe University, Awka 234, Anambra State, Nigeria

Received: September 02, 2011 / Accepted: October 19, 2011 / Published: February 25, 2012.

**Abstract:** Eight flavonoid derivatives: rutin, quercetin–3-glucoside, quercetin, luteolin–7-glucoside, isorhmnetin–3-sulphate, kaempferol–3,7-diglucoside, luteolin and kaempferol have been extracted and characterized from Nipa palm. Mild extraction technique involving the use of HPLC-DAD-MS was used. The structures of the flavonoids were determined on the basis of mass spectroscopy. Separation of the crude extract by paper chromatography (PC) on forestall as solvent system gave one major yellowish brown spot which had  $R_f$  value of 3.9. The  $R_f$  value and maximum absorption from UV spectroscopy were the same as those of quercetin standard. The most prominent compound was quercetin followed by three others: kaempferol–3,7-diglucoside, luteolin–7-glucoside, and isorhmnetin–3-sulphate.

Key words: Nipa palm, flavonoids, extracts, spectroscopy, characterization.

### 1. Introduction

Nypa fruiticans, a mangrove palm, is one of the extra ordinary palms to be seen in the wild state of Australia, where palm is confined to a few small populations on the north-east and north coast [1]. It has also been reported to thrive in other countries especially in brackish waters and Nigeria is one of such [2-3]. Reports have shown that various products are gotten from this palm which include sugar from sap and nuts, feeds for livestock, craft items, alkaloids from the roots, etc. [4-6]. A preliminary work on the palm showed a UV max for flavonoids [7]. Most developing countries of the world especially Nigeria and other West African countries depend on plants for their traditional forms of medicine [8]. Medicinal plants as they are called are used in treating and preventing specific ailment and diseases and as such are considered to play a beneficial

role in health care though based on proofs from western research [8]. It has been stated that hundreds of plant species are recognized as having medicinal values and the medicinal properties being present in one or all the parts of the plant: root, stem, bark, leaf, flower, fruit or seed [9]. It has been discovered that a striking characteristic of plants is that different chemical substances are obtained in members of even the same species in different areas [10]. The development and spread of resistance to the existing antibiotics by microorganisms call for increased efforts in the development of new antibiotics for treatment [9]. Many studies have shown that natural antioxidants from plants sources can effectively inhibit oxidation of food and reduce the risk of age dependent diseases [11]. Extraction of flavonoids from medicinal plants, fruits and vegetables have been studied extensively [12-14] because they present antioxidants with lower toxicity than synthetic antioxidants such as BHA and BHT [15]. New methods of separations have been developed for



<sup>\*</sup>**Corresponding author:** Uche Eunice Ekpunobi, Ph.D., research fields: analytical chemistry, environmental chemistry. E-mail: ask4uche2001@yahoo.com.

extraction of flavonoids [16-19]. In this study, a mild reported method [18] which utilizes a high resolution technique together with DAD chromophore and identification based on UV-visible absorption characterization will be used. This method preserves glycosidic linkages and their combinations with MS facilitate the identification of flavonoids and other glycosides.

# 2. Materials and Methods

#### 2.1 Source of Samples

Nipa palm leaves were collected from the coastal region of Oron, Nipa Palm Utilization Project Centre, National Museum, Oron, Akwa Ibom State, Nigeria.

### 2.2 Separation by Paper Chromatography

1 g of the sample was extracted with 2 mL of HCl and the mixture was heated for 30 minutes at 100 °C. It was filtered and extracted with ethyl acetate, concentrated to dryness and taken up in 2 drops of ethanol. An aliquot was run on a whatman No. 1 filter paper using forestall. Forestall is the mixture of acetic acid-conc HCl-H<sub>2</sub>O in the ratio 30:3:10.

#### 2.3 Extraction

Extraction of flavonoids from Nipa palm leaves (estimated to weigh 0.2-0.8 mg) was done using methanol and water (1:1, v/v) for 2 hrs as reported [18]. The extract was centrifuged to separate particulate matter. The upper layer of solution was removed with a pipette for HPLC-DAD-MS analysis.

#### 2.4 Analysis of Extract

Analysis of the palm extract was performed with an Agilent 1100 liquid chromatography system (HPLC) consisting of an automatic injector, a gradient pump, a HP series 110DAD and an Agilent series 1100 on-line atmospheric pressure elctrospray ionization MS. The flavonoid components were separated on a 2.1-mm-diameter Vydac C4 reversed phase column as previously described [18].

# 3. Results and Discussion

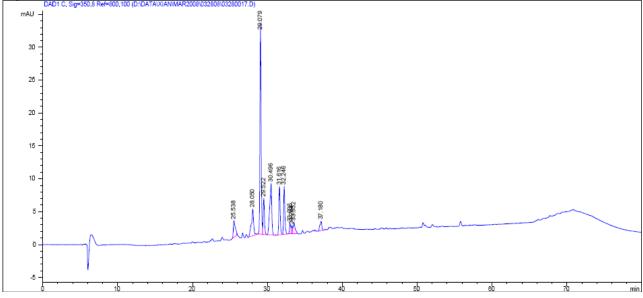
On separation using paper chromatography, a yellowish brown spot was noted on forestall with  $R_f$  value of 39. Comparing this 39 R  $_f \times 100$  value with literature value, it was found to be probably quercetin [19].

The chromatographic profile of Nipa palm leaves is presented in Figs. 1 and 2. The elution profile in Fig. 1 shows eleven peaks observed out of which nine constituents were identified with two unknowns all under the class of flavonoids. The identification of the peaks was possible through the help of authentic samples, mass and UV spectral measurements as reported. The DAD profile of the visible eleven peaks of the elution profile is shown in Fig. 2. Their peaks eluting after 25 minutes are an evidence for classifying them under flavonoids or chalcones which do not absorb at 254 nm. From Table 1, six major peaks were identified as quercetin-4-glucoside (peak 2), quercetin (peak 3). luteolin–7-glucoside (peak 4), isorhmnetin-3-sulphate (peak 5), kaempferol-3,7diglucoside (peak 6) and the sixth (peak 7) as an isomer of peak 1 and 6. Peak 1, a minor peak, was identified as rutin with absorbance of 7.2 and characteristic retention time of 25.538, UV max of 348 and molecular ion of 609 while peaks 10 and 11 as luteolin and kaempferol with corresponding absorbance of 2.6 and 2.5 respectively. From the six major peaks identified, peak 3 (quercetin) gave the highest absorbance of 53.2 followed by peaks 6 (kaempferol-3,7-diglucoside), 4 (luteolin-7-glucoside), 5 (isorhmnetin-3-sulphate), 7 (as an isomer of peak 1and 6) and 2 (quercetin-4-glucoside). All these flavonoids showed diagnostic MS spectral with intense molecular ions of daughter ions at 285 m/z for luteolin and 285 m/z for kaempferol.

Quercetin is a chemical pigment found mostly in the barks of a wide variety of plants which constitutes aglycone of the glycosides rutin and quercetin [20]. It has anti-inflammatory and antioxidant properties hence it has found a number of uses like antihistamine



Nipa leaves extracted in methanol: water=1:1 for 2 hours





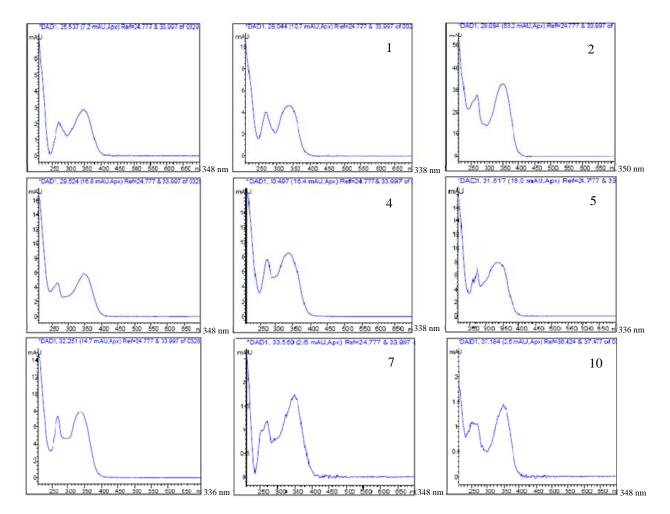


Fig. 2 DAD profile for the identified peaks in Nipa leaves.



Peak No.	Absorbance (mAU)	Retention time (RT) (min)	Max (nm)	[M-H] <sup>-</sup> (m/z)	Fragment ions (m/z)	Likely compound	References
1	7.2	25.538	348	609.1	463, 301, 153	rutin	[21-22]
2	10.7	28.050	338	463	302,153	quercetin-3-glucoside	[18]
3	53.2	29.079	350	301.0	153	quercetin	[18]
4	16.8	29.522	348	447	285	luteolin-7-glucoside	[17, 22]
5	16.4	30.496	338	396	315, 167	isorhmnetin-3-sulphate	[17, 22]
6	18.0	31.616	336	609.1	447, 285, 137	kaempferol-3,7-diglucoside	[21]
7	14.7	32.246	336	609.1	463, 301, 153	isomer of peaks 1 & 6	[21]
8	-	33.036	-	-	-	unknown	-
9	-	33.252	-	-	-	unknown	-
10	2.6	33.252	348	285	-	luteolin	[18, 22]
11	2.5	37.180	348	285	137	kaempferol	[21, 23]

Table 1	Identified compounds in lease	sample of Nipa palm by	HPLC-DAD-MS.

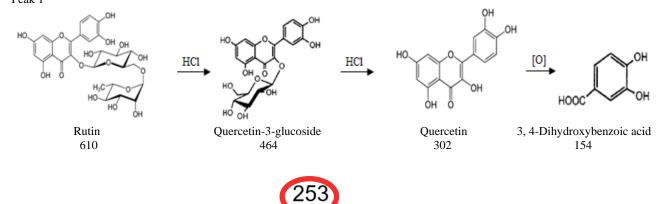
properties [24], helping control allergies and asthma [25], stabilizing small blood vessels [26], protecting against heart attack and strokes [27], helping prevent or treat different types of cancer [28], helping with symptoms of chronic prostatitis [29], relieving some of the neurologic complications of diabetes, etc. [30]. Luteolin was reported to be more effective in antioxidants properties than quercetin [31]. It is a naturally occurring flavonoid which exhibits a wide spectrum of pharmacological properties [32]. It was reported to have anti cancer properties by poisoning eukaryotic DNA topoisomerase I. [33], anti allergic properties and as a potent inhibitor of human mast cell activation through the inhibition of protein kinase C activation and Ca<sup>2+</sup> influx [34]. Flavonoids generally have been reported to possess antioxidants properties; these inhibit cancer cell proliferation in vitro [35], anti viral, anti parasitic, anti cancer activities. A chalcone type extract of UV max 348 nm and molecular mass of 448 and peak 4 has been identified as luteolin-7-glucoside. This compound is one of the most common yellow dyes and occurs in many species of plant dyestuff [20]. Acid hydrolysis also yields Peak 1

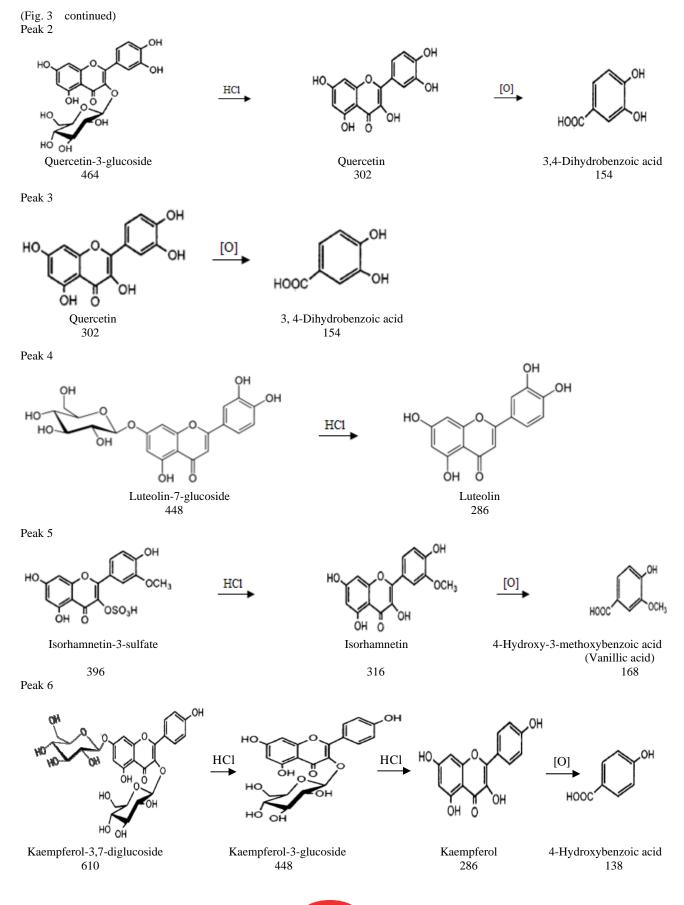
aglycone, luteolin.

Flavonols generally refer to 3-hydroxyflavones of which the most common members are derivatives of quercetin [36]. Quercetin 3-sulfate and isorhamnetin 3-sulfate (M of 382 and 396, respectively) are the most distinctive feature of the flavonol-type dyes [37]. It was reported that the same species of substituted 3-hydroxylflavone which were identified an archaeological samples were same as gotten from F. haumanii specimen [38]. In a pioneering work on natural dyestuff, it was reported that flavonoids dyes have been utilized in textile [39].

#### 4. Possible Mechanism of Fragmentation

The possible mechanism fragmentation is shown in Fig. 3. It was evident that all the aglycones are 3-hydroxy flavones and all seem to have glycosyl or sulphate groups on the 3-hydroxyl group as shown by their UV max which is typical of 3-o-substituted flavones except for peak 4 and 10. By a way of contrast, glycosides of luteolin lack a 3-hydroxyl group and its UV max did not shift when the glycoside was hydrolysed.





105

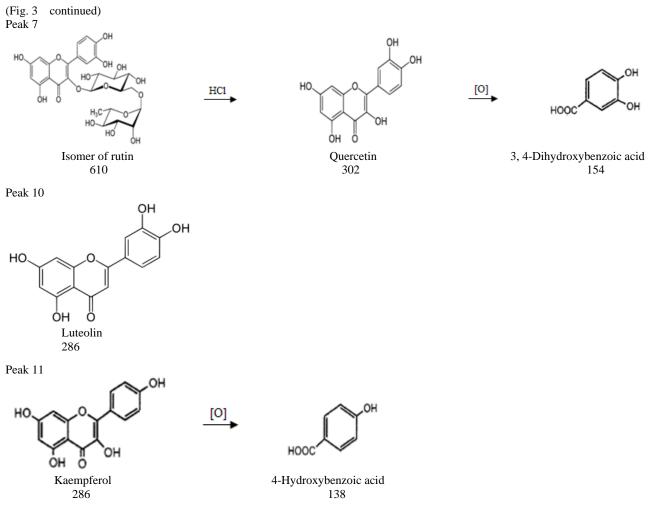


Fig. 3 Schematic representations of fragmentation of the identified peaks 1-7, 10-11.

# 5. Conclusion

The phytochemical results showed that this palm is a good medicinal plant rich in flavonoids. The flavonoids content for instance could be used as antioxidants, anti histamine, and anti-inflammatory properties and are effective against a wide variety of diseases [32]. For instance it has been reported that since ancient times man has utilized alkaloids as medicine. The results have been showed that Nipa palm leaves contain high percentage of flavonoids. The result of the paper chromatography (PC) separation on forestall as solvent system gave one major yellowish brown spot which had  $R_f$  value of 3.9. The  $R_f$  value and maximum absorption from UV spectroscopy were the same as those of quercetin standard. Also from the result of the HPLC-DAD-MS analysis, the most prominent compound was quercetin followed by kaempferol-3,7-diglucoside, luteolin-7-glucoside, and isorhmnetin-3-sulphate. Quercetin in particular has been noted for a number of uses like antihistamine, helping control allergies and asthma, stabilizing small blood vessels, protecting against heart attack and strokes, helping prevent and treat different types of cancer, helping with the symptoms of chronic prostatitis (swelling of the prostrate gland), and relieving some of the neurologic complications of diabetes.

# Acknowledgments

The authors acknowledge Prof. Richard Laursen and group of Department of Chemistry, Boston University,



106

Boston, Massachusetts, USA, for their assistance in the extraction and analysis of extracts as well access to their articles. Jana Sanyova of Institut Royal du Patrimoine Artistique, Belgium, is acknowledged for access to her article. Mr. Aaron Effiong of Nipa Palm Utilization Project Centre, Oron, Akwa-Ibom State, Nigeria is also acknowledged for his assistance in the sample collection and access to their literature collections.

#### References

- John, D.; Robert, T. Notes on the Mangrove Palm, *Nypa fruiticans* Wurmb. *Queensland: & Cycads* 1993, 41, Oct-Dec., 1-5.
- [2] Bilery, F. M. Description of the Queensland Form of *Nypa fruiticans*. *Proceedings of the Royal Security of Queensland* **1888**, *5*(4), 146-148.
- [3] Johnson, D. J. In Palms as Multipurpose Cash and Subsistence Tree Crops, Proceedings of an International Workshop, held in Pataya, Thailand, Nov. 2-5, 1987; pp 222-236.
- [4] Umotong, S. E. In *The Importance of Nipa Palm as a Resource Material for Income Generation Activities*, Proceeding of the Conference of Palm Control by Utilization Project, Oron, Nigeria, 1998; pp 1-5.
- [5] Khieu, B.; Preston, T. R. Conserving Bio-Diversity and the Environment and Improving the Wellbeing of Poor Farmers in Cambodia by Promoting Pig Feeding Systems Using the Juice of Sugar Palm. [Online] 1995, http://www.cipav.org.co/Irrg/Irrd7/2/7.html.
- [6] Hamilton, L. S.; Murphy, D. H. Use and Management of Nipa Palm (*Nypa fruiticans*, areacaeae): A Review. *Economic Botany* 1988, 42(2), 206-213.
- [7] Ekpunobi, U. E.; Eboatu, A. N. Extraction of Sugar from Nipa Palm (*Nypa fruiticans*). J. Chem. Soc. Nig. 2007, 33, 352-354.
- [8] Soejarto, D. D.; Farnsworth, N. R. Global Importance of Medicinal Plants. In *Conservation of Medicinal Plants;* Akerele, O., Heywood, V., Synge, H., Eds.; Cambridge University Press: Cambridge, 1991; pp 231-240.
- [9] Srivatave, J.; Lambert, J.; Vietemeyer, N. Medicinal Plants an Expanding Role in Development. World Bank Technical Paper No. 370: Washington D. C. 20433, USA, 1996; pp 1-2.
- [10] Kharn, M. R.; Ndu alio, G.; Nkunga, M. H. H.; Wever, H.; Sawheny, A. N. Studies on African Medicinal Plants for Antibacterial Activity. *African Journal of Medicinal Sciences* 1985, 15, 65-69.
- [11] Burda, S.; Oleszek, W. Antioxidant and Antiradical

Activities of Flavonoids. *Journal of Agriculture and Food Chemistry* **2001**, *49*, 2774-2779.

- [12] Cai, W.; Gu, X.; Tang, J. Extraction, Purification and Characterization of the Flavonoids from *Opuntia Milpa alta* skin. *Czech. J. Food Sci.* **2010**, *28*(2), 108-116.
- [13] Ding, Z. Studies on Extraction and Isolation of Flavonoids from *Ginkgo* Leaves. *Journal of Food Quality* **1999**, 22(6), 693-700.
- [14] Pekkarinen, S. S.; Heinonen, I. M.; Hopia, A. I. Flavonoid Quercetin, Myrcetin, Kaemferol and (+)-Catechin as Antioxidants in Methyl Linoleate. *Journal of the Science* of Food and Agriculture **1999**, 79, 499-506.
- [15] Andallu, B.; Varadacharyulu, N. C. Antioxidant Role of Mulberry (*Morus indica* L. cv. Anantha) Leaves in Streptozotocin-Diabetic Rats. *Clin. Chim. Acta.* 2003, *338*, 3-10.
- [16] Liu, B.; Zhu, Y. Extraction of Flavonoids from Flavonoid-Rich Parts in Tartary Buckwheat and Identification of the Main Flavonoids. *Journal of Food Engineering* 2005, 78(2), 584-587.
- [17] Sanyova, J.; Reisse, J. Development of a Mild Method for the Extraction of Anthroquinones from Their Aluminium Complexes in Madder Lakes Prior to HPLC Analysis. J. Cul. Heritage 2006, 7, 229-235.
- [18] Zhang, X.; Laursen, R. A. Development of Mild Extraction Method for the Analysis of Natural Dyes in Textiles of Historical Interest Using LC-diode Array Detectors-MS. Anal. Chem. 2005, 77(7), 2022-2025.
- [19] Harborne, J. B. Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis, 3rd ed.; Chapman and Hall: London, 1998; pp 1-36, pp 203-205.
- [20] Zhang, X.; Boytner, R.; Cabrera, J. L.; Laursen, R. Identification of Yellow Dye Types in Pre-Coulumbia Andean Textile. *Anal. Chem.* 2007, 79(4), 1575-1582.
- [21] Kinsella, J. E.; Frankel, E.; German, B.; Kanner, J. Possible Mechanisms for the Protective Role of Antioxidants in Wine and Plant Foods. *Food Technol.* **1993**, *47*, 85-89.
- [22] Hen, X. Q.; Xiao, J. B. RP-HPLC-DAD Determination of Flavonoids: Separation of Quercetin, Luteolin and Apigenin in Marchantia convolula. J. Plant Res. 2005, 4, 175-181.
- [23] Salawu S. O.; Akindahunsi A. A HPLC/DAD/MS Analysis of Flavonoids and Cynamoyl Derivativesin Veronica amygdalina leaves. [Online] 2007, http://publication.ictp.it
- [24] Igilo, G. O.; Wieslaw, L.; Marian, J.; Stanislaw, B.; Fafunso, M.; Fasanmadet, A. A. Flavonoids from *Veronica amygdalina* and Their Antioxidants Activities. J. Agric. Food Chem. **1994**, *42*, 2445-2448.
- [25] De Vries, J. H.; Janssen, P. I.; Hollman, P. C.; Van Staveren, W. A.; Katan, M. B. Consumption of Quercetin



and Kaempferol in Free Living Subjects Eating a Variety of Diets. *Cancer Lett.* **1997**, *114*, 141-144.

- [26] Soliman, F. M.; Shehata, A. H.; Khaleel, A. E.; Ezzat, S. M. An Acylated Kaempferol Glycoside from Flowers of *Freniculum vulgare* and *F. duke. Molecules* 2002, 7, 245-251.
- [27] Harbone, J. B.; William, C. A. Advances in Flavonoids Research Since 1992. *Phytochemistry* 2000, 55, 481-504.
- [28] Hertog, M. G.; Hollman, P. C.; Katan, M. B.; Kromhout, D. Intake of Potentially Anticarcinogenic Flavonoids and their Determination in Adults in the Netherlands. *Nutri. Cancer* 1993, 20, 21-29.
- [29] Tomas-Barberan, F. A.; Gil, M. I.; Ferreres, F.; Tomas-Lorente, F. Flavonoid p-Coumaroyl and 8-Hydroxyflavone Allosyl Glucosides in Some Labiatal. *Phytochemistry* **1992**, *31*, 3087-3102.
- [30] Ray Sahelian, M. D. Natural Treatment for Diabetes Including Herbs and Supplements. Newsletter. 2008. http://www.raysahelian.com
- [31] Braune, A.; Gutschow, M.; Engst, W.; Blaut, M. Degradation of Quercetin & Luteolin by *Eubacterium* ramulu. Applied and Environmental Microbiology 2001, 67(12), 5558-5567.
- [32] Xiao, J. B.; Jiang, X. Y.; Chen, X. Q. Anti Bacterial, Anti-Inflammatory and Diuretic Effect of Flavonoids from *Marchantia Convolula Afr. J. Trad. Comp. Att. Med.* 2005, 12, 244-252.
- [33] Hayatsu, H.; Arimotu, S.; Negishi, T. Dietary Inhibitors of

Mutagenesis and Carcinogenesis. *Mutat. Res.* 1988, 202, 429-446.

- [34] Frankel, E. N.; Kanner, J.; German, J. B.; Parks, E.; Kinsella, J. E. Inhibition of Oxidation of Human Low-Density Lipoprotein by Phenolic Substances in Red Wine. *Lancet* 1993, 341, 454-457.
- [35] Santos, M. R.; Rodriguez-Gomez, M. J.; Justino, G. C.; Charro, N.; Florencio, M. H.; Mira, L. Influence of the Metabolic Profile on the *in vivo* Antioxidant Activity of Quercetin under a Low Dosage Oral Regimen in Rats. *British Journal of Pharmacology* 2008, 153(8), 1750-1761.
- [36] Justesen, U.; Arrigoni, E. Electrospray Ionization Mass Spectrometric Study of Degradation Products of Quercetin, Quercetin-3-Glucoside and Quercetin-3-Rhamno Glucoside Produced by in vitro Fermentation with Human Faecal Flora. *Rapid Commun. Mass Spectrom.* 2001, 15, 477-483.
- [37] Ferreira, E. S. B.; Quye, A.; McNab, H.; Hulme, A. N. Photo-Oxidation Products of Quercetin and Morin as Markers for Characteristics of Natural Flavonoid Yellow Dyes in Anc Textile. *Dyes Hist. Archael.* 2000, 18, 63-72.
- [38] Boytner, R. Andean Textile Tradition; Young-Sanchez, M., Simpson, F. W.; Eds.; Denver Art Museum: Denver Co, 2006; pp 43-74.
- [39] Wouters, J.; Rosario-Chirinos, N. Dye Analysis of Pre-Columbian Peruvian Textiles with High Performance Liquid Chromatography and Diode-Array Detection. J. Am. Inst. Conserv. 1992, 31(2), 237-255.

